

***PENNINGTON BIOMEDICAL RESEARCH CENTER
BATON ROUGE, LOUISIANA
INSTITUTIONAL BIOSAFETY COMMITTEE***

Dr. Diana Coulon
Dr. Prachi Singh

Dr. Timothy Heden
Dr. Abigail Fish (Non-Voting Member)

Dr. Krisztian Stadler, Chair
Dr. Dr. Stefany Primeaux, Vice-Chair

Dr. Leonard Ezell, Community Representative
Jennifer Strickland, Community Representative
Dr. Christopher Morrison, Responsible Official

Dr. Erik Westin
Dr. Sujoy Ghosh

Dr. Christy White
Dr. Sangho Yu

MEETING MINUTES

Thursday, July 9, 2026

2:00 PM – B2001

Institution	Pennington Biomedical Research Center Baton Rouge, LA
Meeting Date	Thursday, July 09th, 2026
Meeting Time	2:02 PM – 2:45 PM
Meeting Type	In person meeting
IBC Members Present	1. Dr. Krisztian Stadler, (Chair) 2. Dr. Stefany Primeaux, (Vice Chair) 3. Dr. Christy White (Safety Officer) 4. Dr. Erik Westin 5. Dr. Timothy Heden 6. Dr. Sangho Yu 7. Dr. Prachi Singh

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	8. Jennifer Strickland (Community Non-affiliated Member)
IBC Members Absent	Abigail Fish (Non-Voting Member) Dr. Diana Coulon (Animal expert) Dr. Sujoy Ghosh Leonard Ezell (Community Non-affiliated Member)
Quorum	Present
Other Individuals in Attendance	N/A
Call to Order	2:02 PM
Conflicts of Interest	<i>The IBC Chair reminded all members present to identify any conflicts of interest as each registration is reviewed.</i>
Review and approval of previous meeting minutes	May 2026 Motion: Dr. Stefany Primeaux Second: Dr. Sangho Yu

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	All in Favor 8/8
Review of Prior Business	N/A

New IBC Registrations and Amendments for Review

PI Name(s)	Prachi Singh, PhD
Registration Number/Title	Registration: R2604 Cellular and molecular mechanisms in cardiometabolic disorders.
Project Overview	The research primarily focuses on mechanisms underlying development of cardiovascular and metabolic disorders such that new therapeutic cardiovascular disease and type 2 diabetes prevention strategies may be developed. Specifically, they explore the interactions between adipose tissue, skeletal muscle, sleep and cardiometabolic disorders using in-vitro and clinical studies. They hypothesize that poor sleep contributes to cellular and molecular alterations that underlie increased cardiometabolic risk among poor sleepers. The primary goal is to identify the cellular and molecular mechanisms that may contribute to metabolic dysfunction. In their

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	clinical research studies, they obtain biospecimens from research volunteers for specific uses and future studies (archived samples). Biospecimens include skeletal muscle, adipose tissue, blood, serum, plasma, saliva, urine, and stool samples. All biospecimens will be collected from research volunteers at PBRC. The lab also uses primary human cell cultures (established from study participants or commercial sources) for their studies to identify causal pathways and cellular consequences. The in-vitro studies in primary human cell cultures also include 1) manipulating protein expression via use of viral vectors to overexpress or silence gene expression; and 2) use of radioactive molecules to examine cellular metabolism.
NIH Guidelines Section	Section III-D-1-a
Risk Assessment and Discussion	1) Human samples: There is a potential risk of exposure to bloodborne pathogens, cuts from sharps, and infectious diseases. All staff receive required biosafety and safety training, use appropriate personal protective equipment (PPE), and follow proper procedures for handling and disposing of samples and sharps. All human samples are treated as potentially infectious. Employees who may be exposed to human fluids are also offered the Hepatitis B vaccine. 2) Viral vectors: The adenovirus and lentivirus used in this research are commercially produced, replication-deficient, and used only in cultured cells under Biosafety Level 2 (BSL-2) conditions. Staff are trained in their safe handling and disposal. 3) Radiation: Small amounts of low-level radioactive materials are used in designated laboratory areas. Staff receive required radiation safety training and follow approved handling and disposal procedures under the supervision of authorized personnel.

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Training	All required training courses are complete for all lab staff listed in the registration.
Occupational Health Representative review (if applicable):	All staff receive bloodborne pathogen training and have received or formally declined the hepatitis B vaccine.
Biosafety Level Assignment	BSL-2
IBC Vote	A motion was made and seconded to approve the registration pending changes or conditions to be met. All in favor 7/7 Conflicts of Interest: Dr. Prachi Singh stepped out of the meeting at 2:07

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Modifications Requested	1. Clearly explain the purpose of the study, including the main research question (hypothesis) and overall goals. 2. Elaborate on where biospecimens are stored. 3. Include all tissue types used in the study, not just adipose and skeletal muscle, in the experimental procedures section. 4. Clarify that human samples should be handled in a biosafety cabinet whenever possible. If any procedures must be performed outside the cabinet, explain why and describe the additional safety precautions and required personal protective equipment (PPE). 5. Clearly define the required PPE for handling human samples and viral materials, including when gloves, lab coats, face masks, face shields, and safety goggles are required. 6. Clarify that liquid biological waste is disinfected using a final bleach concentration of 10% before disposal. 7. Clearly state where chemical fume hoods are used in the study, or remove references to them if they are not needed. 8. Better explain the gene manipulation experiments by identifying which genes are being overexpressed or knocked down, whether the same genes are used in both approaches, whether the methods are combined, and include any missing target genes.
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	9. Identify the species (such as human or mouse) for all biological materials used in the study.
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PI Name(s)	Robert Kesterson, PhD
Registration Number/Title	Registration: R2605 Cancer precision medicine laboratory research.
Project Overview	This project studies neurofibromatosis type 1, also known as NF1, and related genetic conditions that can lead to tumor formation. NF1 is caused by changes in the NF1 gene, which normally helps regulate cell growth. When this gene does not function properly, certain cells may grow abnormally and form tumors, including tumors associated with nerves and other tissues. The overall goal of this project is to better understand how NF1 mutations and related genetic changes contribute to tumor development and to test experimental approaches that may reduce abnormal cell growth. These approaches include studying NF1 mutations in cultured cells and rodent models, testing methods to restore or increase NF1 activity, and evaluating approved or experimental compounds that may affect tumor growth. The central hypothesis is that restoring NF1 function, modifying disease-related gene activity, or targeting pathways affected by NF1 loss will help reduce tumor-related changes in cells and animal models.

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	As part of this research, inclusion of hazardous materials (viruses, human cells, etc.) and murine models will be necessary for the purpose of testing related experimental hypotheses and thus safety practices will be outlined herein.
NIH Guidelines Section	Section III-D-3-a Section III-D-4-a
Risk Assessment and Discussion	1) Bacteria: The E. coli strain used is attenuated and nonpathogenic. 2) Human cell lines and CRISPR/Cas9: work will be performed under BSL-2 conditions. 3) Viral work: The adeno-associated virus is replication-deficient and can be used under BSL-1 conditions. When a helper virus is present, BSL-2 conditions will be used. Lentiviral and retroviral vectors will also be used in BSL-2 conditions. 4) Polymeric nanoparticles: the primary risk is skin, eye, respirator or injection exposure. Small quantities will be used with appropriate engineering controls. 5) Hormone and drug treatments could cause irritation or other effects. Training and engineering controls will reduce the risk. 6) Risk of purified antibody exposure could result in local irritation, sensitization, hypersensitivity or an immune response. The antibody and plasmid DNA are not infectious or self-propagating. 7) Transgenic rodents: exposure risks include animal contact, sharps injury, splashes, aerosol exposure during animal handling and exposure to administered materials. Risks

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	are controlled through animal facility procedures, PEE, sharps precautions, containment, proper labeling and decontamination. Overall, the risk to laboratory staff and the environment is minimized through required training, the use of appropriate protective equipment, engineering controls, proper handling of sharps and laboratory materials, and established safety and waste disposal procedures.
Training	All required training courses are complete for all lab staff listed in the registration.
Occupational Health Representative review (if applicable):	Not all employees have received bloodborne pathogen training or the hepatitis vaccine but these employees will not be working with bloodborne pathogens.
Biosafety Level Assignment	BSL-1 BSL-2, ABSL-1, ABSL-2, BL1-N
IBC Vote	A motion was made and seconded to approve the registration pending changes or conditions to be met. All in favor 7/7 Conflicts of Interest: Dr. Erik Westin stepped out of the meeting at 2:29 pm

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Modifications Requested	1. Clarify and make the laboratory safety procedures consistent throughout the protocol, including the appropriate biosafety level for each type of research and consistent descriptions of animal handling. 2. Update the risk assessment section to include all hazards. 3. Clarfy some statements on waste handling and decontamination. 4. Clarify the hazard training given for work in the animal facility. 5. Note that recapping needles will not be approved unless specifically justified.
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PI Name(s)	Frank Greenway, PhD
Registration Number/Title	Registration: R2606 SCULPT
Project Overview	Many people, especially women, find that fat and cellulite on their thighs are difficult to reduce, even when they lose weight. This study is testing whether a skin cream made from natural compounds found in oranges (naringenin), carrots (beta carotene), and mint (menthol) can help reduce fat in a specific area of the body when applied directly to the skin. The goal of the study is to see whether applying this cream to one thigh

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	twice a day for eight weeks can make that thigh slightly smaller and smoother compared with the other thigh, which will receive a similar looking cream without the active ingredients. Researchers will compare the two thighs within the same participant. The study will also examine whether the cream increases the breakdown of fat cells in the treated thigh. This will be measured using a technique called micro dialysis, which allows researchers to measure substances released when fat is broken down. A sample will be taken from the thigh and micro dialysis performed on it.
NIH Guidelines Section	N/A
Risk Assessment and Discussion	This study involves collecting very small amounts of tissue fluid from healthy adult volunteers using sterile, single use micro dialysis probes. No infectious agents, genetically modified materials, or cell cultures are used. The main risk is the possibility of accidental contact with blood or body fluids during sample collection or handling. This risk is considered low and is minimized by using sterile equipment, protective clothing and gloves, proper sharps disposal, required staff training, and established procedures for safely handling and disposing of human samples.

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Training	All required training courses are complete for all lab staff listed in the registration.
Occupational Health Representative review (if applicable):	All staff receive bloodborne pathogen training and have received or formally declined the hepatitis B vaccine.
Biosafety Level Assignment	BSL-2
IBC Vote	A motion was made and seconded to approve the registration after secondary review by a subcommittee pending changes or conditions to be met. All in favor 8/8 Conflicts of Interest: None
Modifications Requested	1. Clearly explain where all research activities take place by listing every laboratory or building used and briefly describing what work is done in each location. 2. Provide a clearer overview of how human samples are collected, transported, processed, stored, and analyzed, including who handles the samples and the safety measures used during transport and laboratory work. 3. Include the primary risk (human samples) in the risk assessment table. 4. Clearly describe decontamination and disposal procedures.

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	5. The question about training was not answered and needs to be completed. 6. Address the confusion regarding biosafety cabinet and chemical fume hood use. 7. Make PPE use consistent.
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Other Business

	The IBC Chair moved to adjourn the meeting at 2:45 PM. Next Month's Meeting will be scheduled for August 13th at 2 pm.
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