PENNINGTON BIOMEDICAL RESEARCH CENTER
BATON ROUGE, LOUISIANA
INSTITUTIONAL BIOSAFETY COMMITTEE

MEETING MINUTES

Thursday, May14, 2026
2:00 PM – B2001

Institution	Pennington Biomedical Research Center Baton Rouge, LA
Meeting Date	Thursday, May14th, 2026
Meeting Time	2:02 PM – 2:20 PM
Meeting Type	In person meeting
IBC Members Present	1. Dr. Jason Collier, (Chair) 2. Dr. Krisztian Stadler, (Vice Chair) 3. Dr. Christy White (Safety Officer) 4. Dr. Diana Coulon (Animal expert) 5. Dr. Stefany Primeaux 6. Dr. Erik Westin 7. Dr. Timothy Heden

	8. Dr. Sangho Yu 9. Dr. Prachi Singh 10. Leonard Ezell (Community Non-affiliated Member) 11. Jennifer Strickland (Community Non-affiliated Member)
IBC Members Absent	1. Abigail Fish (Non-Voting Member)
Quorum	Present
Other Individuals in Attendance	N/A
Call to Order	2:02 PM
Conflicts of Interest	*The IBC Chair reminded all members present to identify any conflicts of interest as each registration is reviewed.*
Review and approval of previous meeting minutes	March 2026 Motion: Dr. Sangho Yu

	Second: Dr. Stefany Primeaux All in Favor 11/11
Review of Prior Business	Waiting for modifications to be completed by Kesterson Lab for R2602.

New IBC Registrations and Amendments for Review	
PI Name(s)	Stephania Cormier PhD
Registration Number/Title	Registration Renewal: R2603 Immunopathogenesis of viral respiratory pathogens
Project Overview	Human clinical samples will be used to study immune responses to viral respiratory pathogens including respiratory syncytial virus (RSV), influenza (IAV), and SARS-CoV-2. These samples include nasal/tracheal aspirates, nasal swabs, and whole blood and will be supplied from a clinical lab. The lab will analyze these samples for viral pathogens including viral load, cytokines, cellular (flow cytometry), and antibody responses. To understand mechanism, a variety of different transgenic/knockout/wild-type mice will be

	infected with RSV/IAV and lung cells are isolated for the following analyses: cytology (fixed cells), histology (fixed tissue), and flow cytometry/sorting (fixed cells). For these studies RSV/IAV will be produced in the lab. Fluorescently labeled RSV will also be used. In addition, the lab studies the respiratory health effects of combustion derived particulate matter (CDPM). In this project the lab exposes mice or human airway epithelial cells to CDPM. To understand the mechanism, mice/cells are exposed to aerosolized CDPM and lung cells are isolated for the following analyses: cytology (fixed cells), histology (fixed tissue), and flow cytometry/sorting (fixed cells). The goals of this project are to understand how this exposure affects the lungs differently in males and females, identify the biological processes that make males more susceptible, and test potential treatments that could reduce lung damage. Finally, the lab collaborates on a research project to understand the respiratory impacts of e-cigarettes by characterizing the immune cells in the placenta and umbilical tissue. Specifically, the lab will isolate single cells and perform flow cytometry to characterize immune cells using the procedures described in the current protocol.
NIH Guidelines Section	Section III-D-4-a

Risk Assessment and Discussion	1) RSV/IAV can be contracted through the ocular route and by inhalation while performing infections and surgeries. The CDC recommends Standard Precautions within a BSL2 facility with proper PPE. Risk to healthy personnel working with RSV and IAV (A/PR/8/34; mouse adapted strain) is minimal. Personnel are encouraged but not required to receive the IAV vaccine annually. Risk will be minimized through training, engineering controls and proper PPE use. 2) Working with whole human blood and tissues and human cell lines: Risks involved while working with human blood, tissues and cell lines include inhalation of blood borne pathogens via aerosolized particles and via dermal cuts/abrasions. To minimize any such risk, all procedures are performed in a class II biosafety cabinet with proper PPE after proper training. 3) Exposure studies: Exposure of animals to combustion-derived particulate matter (PM) pose a risk of inhalation of aerosols. Risk will be minimized through training, engineering controls and proper PPE use. 4) Working with SARS-CoV-2: SARS-CoV-2 can be transmitted through inhalation of respiratory droplet aerosols and fecal-oral route. Working with human samples and routine diagnostic testing of potential SARS-CoV-2 containing specimens, according to the CDC, can be handled in a BSL-2 laboratory using Standard Precautions and proper PPE.
Training	All required training courses are complete for all lab staff listed in the registration.

Occupational Health Representative review (if applicable):	One employee is in need of bloodborne pathogen training and is signed up for the next scheduled class. She has also been assigned the hepatitis B vaccine compliance.
Biosafety Level Assignment	BSL-2, ABSL-2
IBC Vote	A motion was made and seconded to approve the registration pending changes or conditions to be met. All in favor 11/11 Conflicts of Interest: None
Modifications Requested	1. Tamoxifen does not need to be included in this registration because it is already a well-known chemical with an existing PBRC SOP. Remove all mention of it. 2. In the summary section, include information about combustion-derived particulate matter and human placental and umbilical tissues (added in last year's amendment). 3. In the experimental procedures section, clarify whether rRSV is being given to mice. Also include the work involving human placental and umbilical tissues. 4. Clarify if employees are required to take the influenza vaccine. 5. Include a risk assessment and safety practices for working with human tissues and cell lines. Also add them to the human tissues section. 6. Specify whether samples will be transported using a personal vehicle or a Pennington vehicle.

	7. Correct the NIH guidelines section. 8. Clarify how long influenza virus is shed in mice. 9. Clarify that 10% bleach is used for spill cleanup in the lab but that CBC no longer uses or provides bleach for the animal facility. 10. Update the instructions for what to do if a transgenic rodent escapes so they match the current Escaped Rodent SOP from the CBC. 11. Update the SOPs to clarify that the lab will be handling mouse cages during the infectious period.
Other Business	
	Committee changes: Dr. Collier will be stepping down as chair and Dr. Stadler will be the new chair of the IBC committee. Dr. Primeaux will move to the position as Vice Chair. Dr. Sujoy Ghosh will replace Dr. Collier on the committee.
	The IBC Chair moved to adjourn the meeting at 2:20 PM. Committee will be reviewing the Viral SOP in June. Next Month's Meeting will be scheduled for June 11th at 2 pm.